Selective Binding of Dicarboxylate Substrates by Ditopic Polyammonium Macrocycles

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Three macrocyclic hexaamines 1, 2, and 4 and the acyclic tetraamine 5 and hexaamine 6 have been synthesized. The hexaamines 1, 2, and 4 are ditopic coreceptor molecules containing two triamine subunits which may bind anionic substrates when protonated. The stability constants of the complexes between the protonated forms of the macrocyclic polyamines and terminal dicarboxylates $^{-}O_2C^{-}(CH_2)_m^{-}CO_2^{-}$ as well as amino-acid and dipeptide dicarboxylates have been determined by pH-metric measurements. Around neutral pH, 1 and 2 give mainly complexes of the fully protonated species $1 \cdot 6H^+$ and $2 \cdot 6H^+$, whereas 4 yields predominantly complexes of $4 \cdot 5H^+$ and $4 \cdot 4H^+$. The stability sequences of the complexes formed indicate preferential binding of the dianionic substrates whose length is compatible with the separation of the triammonium binding subunits in the protonated species 1 and 4 and the separation of the corcecptor and the terminal carboxylates of the substrate of complementary length. The complexes of the acyclic ligands 5 and 6 are much weaker and much less selective, indicating a marked macrocyclic effect on both stability and selectivity of binding, *i.e.* on recognition.

Introduction. – The incorporation of two or more binding subunits into a macropolycyclic structure yields coreceptor molecules in which the subunits may cooperate for the multiple binding of a polyfunctional substrate or for the simultaneous complexation of two or more substrates. Such coreceptors add a new dimension to the chemistry of molecular receptors [1]. Thus, cylindrical macropolycycles form selective cryptates of ${}^{+}H_{3}N-(CH_{2})_{n}-NH_{3}{}^{+}$ cations, bound by each terminal $NH_{3}{}^{+}$ group to a macrocyclic subunit, with a stability and selectivity depending on the complementarity between the length of the substrate and the cavity size of the receptor [1–3].

In a similar fashion, strong and selective binding of a dianionic molecule requires the design of a ditopic molecular coreceptor possessing two binding subunits arranged in such a way that they cooperate for substrate binding by interacting with the anionic groups of the target species. The binding subunits themselves must contain a suitable array of interaction sites capable of forming intermolecular bonds to the anionic sites of the substrate.

Macrocyclic [4–8] and macropolycyclic [9–14] polyammonium molecules have been shown to complex strongly and selectively a variety of inorganic and organic anions, thus laying the bases for the developing field of anion-coordination chemistry [1] [10] [13] [15] [16]. The anion-complexation units of these receptor molecules consist of several positively charged binding sites arranged around a cavity defined by the macropolycyclic architecture.

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We have incorporated such polyfunctional binding features in the design of ditopic coreceptors for dianionic substrates. We describe here the synthesis of ditopic macrocyclic polyamines and their binding properties towards terminal dicarboxylate substrates $^{-}O_2C-R-CO_2^{-2}$).

Design of the Polyaza-macrocycles. – In order to achieve the binding of terminal dicarboxylates, the receptor must present two binding subunits located at the poles of the structure and held by the molecular framework. Since electrostatic charge-charge interactions play a dominant role in binding of anions, polyammonium cations should be suitable complexing agents; strongest binding occurs usually with the fully protonated form of the polyamine.

Because most organic anions, especially polycarboxylates, correspond with weak acids and thus are protonated below a pH of *ca*. 4, the receptor molecule should be designed so as to fulfill a compromise between the pH values required for full protonation of the polyamine receptor and the need to remain in a pH range where the substrate to be bound may exist as dicarboxylate. In order to accumulate the highest charge density in each binding subunit of the ligand, the number of ammonium binding sites may be increased, but this will also lower the pH for full protonation. Thus, in the design of a suitable ditopic coreceptor, the following parameters must be taken into account: the number of ammonium binding sites forming the binding subunits; the distance separating the ammonium sites from each other within a binding subunit, and the distance separating the two binding subunits. Structure A gives a schematic representation of the binding of a terminal dianionic substrate to a ditopic coreceptor containing two triammonium subunits located at the poles of the molecule.



²) Preliminary report, see [7].

'Diethylenetriamine' (= dien = 3-azapentane-1,5-diamine) and 'di(trimethylene)triamine' (= dpen = 4-azaheptane-1,7-diamine) are attractive binding subunits, since their lowest pK_a values are 4.25 [17] and 7.72 [18], respectively. For the naturally occurring acyclic polyamines putrescine, cadaverine, spermidine, and spermine, the pK_a values for full protonation are rather high (> 7.5) [19]. These polyammonium cations display weak binding of biologically relevant anions [20]. On the other hand, unprotonated diethylenetriamine and di(trimethylene)triamine groups bind transition metal ions so that macrocycles containing two such units should also be able to form dinuclear cation complexes which may have interesting structures and properties [21].

These considerations on potential binding ability towards both metal cations and anions led to the choice of the macrocycles 1-3. Compounds 1 and 2 are 32- and 38-membered macrocyclic polyamines containing two 'di(trimethylene)triamine' groups as subunits separated by linear chains of 7 or 10 methylene groups. They may also be considered to incorporate two fragments reminiscent of the natural acyclic polyamines cited above which play an important biological role [22]. The 32-membered macrocyclic polyamines **3** is based on 'diethylenetriamine' groups as the binding units separated by 9 methylene groups. The polyamines **5** and **6** may be considered as open-chain reference compounds for comparison with the corresponding macrocycle **2**.

Synthesis of the Macrocycles 1–3 and the Linear Polyamines 5 and 6. – For the macrocyclic polyamines 1–3, the key step, the cyclisation, was achieved by condensation of an α, ω -di(*p*-toluenesulfonamide) with an α, ω -ditosylate in dimethylformamide (DMF) in the presence of caesium carbonate (Cs₂CO₃) following the methods described for the synthesis of diazacyclodecadiene [23] and of polyhetero macrocycles [24].

Macrocycles 32 $\langle N_6-3_2, 7.3_2, 7$ -*Coronand*-6 \rangle [32]*ane*- $N_6(C_7)$ **1** *and* 38 $\langle N_6-3_2, 10.3_2, 10$ -Caronand-6 > [38] ane- $N_6(C_{10})$ 2 (see Scheme 1). The synthesis of 1 and 2 follows a reaction sequence similar to that used earlier for the related macrocycle 4 [4] [25]. The ditosyl derivatives 7 and 14 were obtained in high yield by the tosylation of the corresponding diamines. These two compounds were common starting materials for the two chains used in the cyclization steps. Treatment of 7 and 14 with acrylonitrile in DMF in the presence of K_2CO_3 gave the dinitriles 8 and 15, respectively. Reduction of 8 and 15 with B_2H_6 [26] led to the diamines 9 and 16, which were converted into their tetratosyl derivatives 10 and 17, respectively. On the other hand, treatment of 7 and 14 with 3-chloro-1-propanol in DMF in the presence of excess solid K₂CO₃ gave the diols 11 and 18, which were converted into the di(mesyloxy) derivatives 12 and 19 [27], respectively. Condensation of 12 and 19 with the second cyclization partner 10 and 17, respectively, at 90° in DMF in the presence of $C_{2}CO_{1}$ [28] led to the hexatosyl macrocycles 13 and 20, respectively, which were purified carefully. Removal of the tosyl groups was achieved by treatment with 33% HBr/AcOH/phenol at 80° [23] [29] yielding 1 · 6 HBr or 2 · 6 HBr. This method gave higher yields than the treatment with conc. H_2SO_4 at 100° for an extended period of time [24] [30]. The free macrocyclic hexaamines 1 and 2 were obtained by passing 1.6 HBr or 2.6 HBr over a *Dowex* 1×8 resin in its basic form; they should be stored under N_2 or kept as their hexaammonium salts.

The linear tetraamine 5 was obtained from 17 by removal of the tosyl groups as described above. The linear hexaamine 6 was prepared by condensation of the disodium salt of N,N',4-tri(*p*-tolylsulfonyl)-4-azaheptanediamine, prepared as described earlier





Macrocycle [32]ane-N₆(C₉) **3** (see *Scheme 2*). The synthesis of **3** follows a reaction sequence similar to that used earlier for the synthesis of smaller macrocyclic hexaamines [31]. The monoprotection of 1,9-nonanediol was achieved by treatment with dihydropyran [32] yielding **25** which was converted into the mesyloxy derivative **26**. Condensation of the latter with the disodium salt **24**, prepared as described earlier [33], gave **27** which was refluxed in EtOH/H₂O in the presence of *p*-toluenesulfonic acid [34] yielding the diol **28**. The latter was converted into the di(mesyloxy) derivative **29**. The cyclisation step was performed by condensation of **29** with the tritosyl derivative **23** in DMF in the presence of Cs₂CO₃ yielding the hexatosyl macrocycle **30**. Deprotection of the latter was performed as described above and afforded the hexaamine **3**.



Results. – Protonation Features of the Macrocyclic Polyamines 1–4 and of the Linear Tetraamine 5 and Hexaamine 6. The protonation constants log K_n (= pK_a values) corresponding to the equilibria of the polyamines L = 1–6 (Eqn. 1 and 2) are listed in Table 1. They lead to the distribution curves of the various species represented in Fig. 1 in the case of L = 1 and 3.

$$\mathbf{H}_{n} \mathbf{L}^{(n-1)+} + \mathbf{H}^{+} \rightleftharpoons \mathbf{H}_{n} \mathbf{L}^{n+} \tag{1}$$

$$K_{n} = \frac{[\mathbf{H}_{n}\mathbf{L}^{n+}]}{[\mathbf{H}_{n-1}\mathbf{L}^{(n-1)+}][\mathbf{H}^{+}]}$$
(2)

It may first be noted that in the case of macrocyclic polyamines 1 and 2, the lowest $\log K_n$ values are close to 7, so that they are fully protonated at pH values close to neutrality. The lowest $\log K_n$ value for the compound 3 is around 3.5; thus, at pH ca. 7, the most abundant species in solution is the tetraprotonated form $3 \cdot 4H^+$ (Fig. 1). This clearly shows the importance of the choice of the binding subunits. Furthermore, since the polyammonium forms of 1–6 are anion receptors, the log K_n values found depend on the anion, being higher the stronger the interaction with the anion present in the supporting electrolyte. The latter was chosen so as to minimize such effects of the medium. However, it is clear that in view of this, the pK_a values determined (as well as the stability constants,

n	1	2	3	4	5	6				
1	10.70 (10.85)	> 10.25 (> 10.50) ^b)	> 9.70 ^b)	10.45 (10.50)	10.75	10.80				
2	10.70 (10.60)	$> 10.25 (> 10.50)^{b}$	> 9.65 ^b)	10.35 (10.20)	10.70	10.75				
3	9.85 (9.80)	10.10 (10.15)	9.60	9.05 (9.25)	9.30	9.95				
4	9.60 (9.05)	9.60 (9.45)	9.25	7.90 (8.00)	8.75	9.60				
5	7.90 (7.40)	7.95 (7.65)	4.15	7.15 (7.05)	-	8.05				
6	7.30 (6.65)	7.30 (6.95)	3.55	6.60 (6.40)	- <i>i</i> .	7.45				

Table 1. Protonation Equilibrium Constants log K_n (= pK_a) of the Macrocyclic Polyamines 1-4 and of the Linear Polyamines 5 and 6^a)

^a) In H₂O, at 25°, see Eqn. 1 and 2 for definition of K_n ; supporting electrolyte 0.1M or 0.01M (values in parentheses) Me₄NCl; see also [7] for 1 and 2 and [4] [25] for 4.

^b) The pK_1 and pK_2 values cannot be determined since these compounds are not soluble in H_2O in their unprotonated form.



Fig. 1. Distribution curves of the unprotonated and protonated forms of the macrocyclic polyamines as a function of pH; a) compound 1, b) compound 3. L: unprotonated species; the numbers 1-6 refer to the successive protonated species bearing 1-6 protons; Σ: summation over all the protonated species. In the case of 3, the unprotonated macrocycle L precipitates and is not represented; Σ does not contain LH⁺.

see below) hold specifically for the medium used here. Although complexation of chloride is probably weak, it nevertheless affects the data so that the protonation constants determined in its presence are apparent values. The fact that the three lowest log K_n values of the free ligands 1–2 are appreciably more basic in 0.1M than in 0.01M Me₄NCl, whereas the three highest ones are almost unaffected, is indication of weak chloride binding, as also seen by ³⁵Cl-NMR studies of the chloride resonance [35]. The log K_n values for compound 4 in 0.1M sodium *p*-toluenesulfonate (TsONa) also show the same effect [25]. The pK_a values of the anionic substrates have also been determined under the same conditions for use in the stability-constant computations.

Complexation Features of the Macrocylic and Linear Polyamines 1–6. The stability constants $\log K_s^n$ corresponding to the equilibria of the polyammonium ions $H_n L^{n+}$ (L = 1–6) with various dicarboxylate anions A^{m-} (Eqn.3 and 4) have been determined by pH-metric titration (see *Exper. Part*); they are listed in *Table 2*. Those for 4 are taken from earlier work [4].

$$\mathbf{L} + n\mathbf{H}^{+} + \mathbf{A}^{m-} \rightleftharpoons (\mathbf{H}_{n}\mathbf{L}\mathbf{A})^{(n-m)+}$$
(3)

$$\mathbf{K}_{s}^{n} = \frac{[(\mathbf{H}_{n}\mathbf{L}\mathbf{A})^{(n-m)+}]}{[\mathbf{H}^{+}]^{n}[\mathbf{L}][\mathbf{A}^{m-}]}$$
(4)

Dicarboxylate anions	n^{c})	Macrocyclic and linear polyamines								
(<i>m</i>) ^b)		1	2	3	4	6 ^d)				
Oxalate ^b) (0)	6	(3.20)	(6.30)		(3.80)					
	5	(2.50)	(4.70)		(3.20)					
	4	(1.90)	(2.85)		(2.60)					
Malonate ^b) (1)	6	(2.75) 3.80	(3.80) 4.05		(3.30)					
	5	(2.05) 2.90	(2.65) 3.05		(2.60)					
	4	(1.35) 1.50	(2.20) 1.95		(2.45)					
Succinate ^b) (2)	6	(3.40) 4.30	(3.0) 3.15		(2.40)					
	5	(2.85) 3.30	(2.35) 2.40	(3.65)	(2.05)					
	4	(2.45) 2.55	(2.20) < 1.2	(1.20)	(1.80)					
Glutarate ^b) (3)	6	(3.40) 4.40	(2.90) 3.30	(6.10)	(2.35)	(1.95)				
	5	(2.90) 3.40	(2.45) 2.55	(5.50)	(2.30)	(1.65)				
	4 ^d)	(2.50) 2.80	(2.40) 1.55	(2.95)	(2.20)	. ,				
Adipate ^b) (4)	6	(2.30) 3.20	(2.95) 3.20	(4.50)	(2.35)	(2.05)				
1 / (/)	5	(1.90) 2.65	(2.50) 2.55	(3.80)	(2.30)	(1.80)				
	4 ^d)	(1.65) 1.75	(2.40) 1.45	(1.54)	(2.20)	()				
Pimelate (5)	6	(2.25) 3.10	(3.40) 4.40			(2.25)				
(-)	5	(1.85) 2.40	(2.85) 3.55	(3.10)		(2.0)				
	4	(1.85) 1.60	(2.70) 2.75	(1.10)	(2.00)	()				
Suberate ^b) (6)	6		(3.45) 4.25							
	5		(3.00) 3.45							
	4		(2.65) 2.65							
Azelate ^b) (7)	6	_	(3.20) 3.60	_						
,	5	-	(2.85) 3.15	_		-				
	4	_	(2.55) 2.50		-					
Sebacate ^b) (8)	6	_	(3.05) 3.50		_	_				
	5	_	(2.90) 3.15	-		· _				
	4	_	(2.65) 2.40			-				
Maleate ^b) (2)	6	4.30	_		(3.70)	_				
	5	3.30			(2.95)					
	4	2.30		-	(2.70)	-				
Fumarate ^b) (2)	6	4.10	-	_	(2.20)	-				
	5	3.25		_	(1.90)	_				
	4	2.50	-		(1.75)	-				
N-Acetyl-L-aspartateb) (2)	6	4.10	3.35		-	-				
	5	3.10	2.60		-					
	4	2.30	< 2		-					
N-Acetyl-L-glutamate ^b) (3)	6	4.15	3.25			-				
	5	3.10	2.60	-	-	-				
	4	2.30	< 2	_	_	-				
N-Acetyl-L-(1-glutamyl)-	6	3.15	4.30	-	-	-				
glycinate ^b) (6)	5	2.40	3.50	-	-	-				
	4	< 2	2.40	_	-	-				

Table 2.	Stability	Constant	log	Ks	(±0.2), f	for	Dicarboxylate-Anio	n	Binding	by	the	Polyammonium	Receptor
Molecules 1–6 in Aqueous Solution ^a)													

^a) The log K_s values are determined in the presence of either 0.01M or 0.1M (values in parentheses) Me₄NCl.

Þ) Chain length in $^{-}O_2C-(CH_2)_m-CO_2^{-}$ or number of atoms separating the two terminal carboxylate groups.

, c) d) Number of protons involved in complexes of the type (receptor, anion, $n H^+$).

For compound 5 with glutarate and adipate, $\log K_s = (1.60)$ and (1.80), respectively.



Fig. 2. Distribution curves for the species present in the medium as a function of pH in two cases; a) macrocyclic polyamine 1, and b) macrocyclic polyamine 3 in presence of glutarate ${}^{-}O_2C - (CH_2)_3 - CO_2^{-}$. C4, C5, C6; complexes (L, A²⁻, nH⁺) with n = 4,5,6. Σ C: summation over all complexes. Σ L: summation over all uncomplexed ligand species (except unprotonated and monoprotonated for 3).

The results lead to the distribution curves of the various species present in solution; two examples are shown in *Fig.2. Fig.3–5* (see below) give graphical representations of the stability constants in comparative series of complexes in order to provide a clearer visualization of the selectivity sequences observed.

The polyamines 1–6 were used as their hydrochloride salts. The supporting electrolyte was 0.1M or 0.01M Me₄NCl. In the latter case, the ionic strength is only approximately constant over the titration, so the absolute K_s^n values are less reliable than the relative ones. As mentioned above, weak complexation of chloride competes to some extent with the anion studied; consequently the stability constants determined are apparent constants, the real values for a given anion being even higher than those listed in *Table 2*. The log K_s^n values are appreciably higher in 0.01M than in 0.1M Me₄NCl; this may be due both to the known marked decrease in binding constant when the ionic strength increases and to chloride competition.

The stoichiometry of the complexes was assumed to be 1:1 in the data-analysis procedure. However, the presence of a certain amount of higher-order complexes cannot be excluded. This holds especially for the smaller substrates (oxalate, malonate) for which simultaneous binding of a single molecule to the two binding subunits of the receptor is geometrically unfeasible, unless there is a marked deformation of the macrocycle; the high stability constants calculated for oxalate and malonate, assuming 1:1 stoichiometry, probably result from the presence of 2:1 dianion/hexacation complexes (see *Table 2*).

Discussion. – The synthetic procedures developed here provide a general route to ditopic macrocyclic polyamines containing two chelating subunits separated by structural units of given geometrical features.

The distribution of protonated species, as illustrated in *Fig. 1*, substantiates the choice of 'di(trimethylene)triamine' subunits in order to obtain receptor molecules which are fully protonated around neutral pH, as it is the case for 1 and 2. Comparison with 4 [4] [25] also shows that increasing the separation of the subunits markedly increases the pK_a 's for full protonation, as expected. On the other hand, 3 which contains two 'diethylene-triamine' subunits is only tetraprotonated in the same pH domain. This has important consequences for the nature, stability, and distribution of the complexes formed.

As a result, only 1 and 2 are expected to yield preferentially the most stable complexes, which are those of the hexaprotonated forms. This is clearly seen in *Fig. 2* which compares complexation of glutarate by 1 and by 3. Thus, the binding properties of 1 and 2 around neutral pH are mainly those of 1.6H^+ and of 2.6H^+ forming the $(L \cdot 6 \text{H}^+, \text{A}^{2-})$ complexes. On the other hand, the analogous complexes of 3 have very low abundance, the species formed being of the type $(3.5 \text{H}^+, \text{A}^{2-})$ and $(3.4 \text{H}^+, \text{A}^{2-})$.



Fig. 3. Graphical representation of the stability constants log K_s of the complexes formed by the polyammonium macrocycles $1.6H^+$ (\bigcirc) and $2.6H^+$ (\bigcirc) with the dicarboxylates $-O_2C - (CH_2)_m - CO_2^-$ as a function of chain length m. See also Table 2 and text.



Fig. 4. Graphical representation of the stability constants log K_s of the complexes formed by the polyammonium macrocycle 3 with the dicarboxylates $^{-}O_2C - (CH_2)_m - CO_2^{-}$ as a function of chain length m. C4, C5 and C6 correspond to the $(3, A^{2-}, nH^+)$ species with n = 4,5, and 6. See also Table 2.

The polyammonium macrocycles 1-4 are efficient *anion-receptor molecules* forming strong complexes with dicarboxylate anions in aqueous solution (*Table 2*). The most stable species are those of the hexaprotonated forms $L \cdot 6H^+$ which exert the largest electrostatic interactions. The stability decreases markedly for the ligands of lower protonation state.

The *complexation selectivity* presents a marked structural dependence. Each receptor 1-3 shows a selectivity peak among the homologous $^{-}O_2C - (CH_2)_m - CO_2^{-}$ substrates (Fig. 3 and 4). Furthermore, the selectivity peak shifts from m = 2 and 3 to m = 5 and 6 on going from 1 to 2, which corresponds to the same increase in length (by three CH_2) groups) both for the most strongly bound dicarboxylates and for the $(CH_2)_n$ bridges separating the two binding subunits in 1 (n = 7) and in 2 (n = 10). The smaller receptor 4 (n = 3) binds best the shorter dicarboxylates. There is, thus, a close correspondence between binding-site separation and substrate length. The low selectivity between the two neighbouring substrates m = 2 and 3 for 1 and m = 5 and 6 for 2 (Fig. 3) may result from the opposite effects of stronger electrostatic forces in the shorter substrate (m = 2 or 5) and better structural fit of the longer one (m = 3 or 6), as well as from unsufficient rigidity of the present receptors. On the other hand, a definite selectivity peak is found for the binding of glutarate by protonated 3 (Fig. 4). This may be due to the higher structural localisation of the ammonium sites when the subunits are of the dien rather of the dpen type; the same factor may explain the strong binding shown by 3 as compared to 1 and 2 in the same protonation state.



Fig. 5. Graphical representation of the stability constants log K_s of the complexes formed by the polyammonium macrocycles $1 \cdot 6 H^+$ (\bigcirc) and $2 \cdot 6 H^+$ (\square) with amino-acid and depeptide dicarboxylates as a function of the number of atoms m separating the two $-CO_2^-$ groups. See also Table 2.

Selective binding of *biological dicarboxylate substrates* also occurs following the same lines. Thus, 1 binds preferentially N-acetyl-L-aspartate and N-acetyl-L-glutamate with respect to the dipeptide N-acetyl-L-(1-glutamyl)glycinate, whereas the reverse holds for 2, in line with the chain length of these substrates (*Fig. 5*).

Comparison of the complexation ability of compounds 2 and 6 demonstrates a pronounced *macrocyclic effect*. The linear hexaamine 6 forms much less stable complexes than its macrocyclic analogue 2 and shows almost no selectivity peak (*Table 2*).

Both the stability and the selectivity of the complexes formed by the macrocycles 1–3 display the operation of special *structural features*. The high stabilities observed for the optimal coreceptor-dicarboxylate pairs result from the incorporation of two binding subunits in the macrocycle and from double (ditopic) carboxylate-group/triammonium-site binding; this is indicated by the low stabilities found for the single-site interactions of butyrate with $2 \cdot 6 \,\mathrm{H}^+$ and of pimelate or butyrate with the subunit reference triammonium cation ${}^{+}\mathrm{H}_3\mathrm{N}-(\mathrm{CH}_2)_3-\mathrm{NH}_2^+-(\mathrm{CH}_2)_3-\mathrm{NH}_3^+$ (log $K_i^* < 2.0$).

The peak selectivity observed as a function of chain length reveals a dominant structural factor in dicarboxylate binding. Electrostatic charge-charge interactions which favor binding of anions of high charge density were usually found to dominate both the strength and the selectivity of complexation between highly charged partners [4] [6]. However, an effect of ring-size was observed earlier for ligands of type 4 [4]. The present chain-length selection describes a linear molecular recognition process analogous to that found for diammonium substrates [1–3]. It may be attributed to structural complementarity between the dianionic substrate and the ditopic coreceptor molecules $1 \cdot n H^+$, $2 \cdot n H^+$, and $3 \cdot n H^+$ in which the two binding subunits cooperate for substrate binding. The terminal anionic groups of the dicarboxylate would each interact with a di- or triammonium unit of the coreceptor, the polymethylene-chain stretching between the polymethylene bridges of the macrocycle, in more or less extended conformations (see also structure A). Highest stability of the complex should correspond to the best fit between substrate length and site separation in the coreceptor; substrates that are either too short or too long form less stable complexes. Linkage of the binding subunits by more rigid bridges than polymethylene chains may be expected to increase the complexation selectivity, *i.e.* to improve the recognition of molecular length operated on the dianionic substrates.

Conclusion. – The present results on the binding of terminal dicarboxylate anions, together with those obtained earlier for the binding of diammonium cations of different chain lengths by macrotricyclic receptor molecules, demonstrate that it is possible to design coreceptor molecules for the selective ditopic binding of difunctional molecular substrates [1–3]. Higher stabilities and selectivities may be achievable by incorporating more rigid structural unit and by designing amphiphilic anion receptors of speleand type [1]. Extension to polytopic receptors may follow similar lines and lead to more elaborate molecular recognition *via* polyfunctional binding. Furthermore, subunit cooperation in coreceptor molecules containing suitable structural elements may allow to perform cocatalysis and cotransport processes [1] [36] [37].

Experimental Part

1. General. M.p.: uncorrected. pH-Metric measurements: Metrohm-636 titrimeter; the cell was thermostated at 25° ± 0.1°, the soln. stirred, and all measurements were performed under N₂. The log K_n values of the compounds were determined by titration with 0.1N NaOH of a soln. containing typically 10⁻³M of the polyammonium salt in the presence of Me₄NCl (0.1M or 0.01M). The log K_s values of the complexes were determined by titration with 0.1N NaOH of a soln. containing to polyamine and 5×10^{-3} M of the desired dianions in the presence of Me₄NCl (0.1M or 0.01M). The log K_s values of the complexes were determined by titration with 0.1N NaOH of a soln. containing 10⁻³M of the HCl salt of the desired polyamine and 5×10^{-3} M of the desired dianions in the presence of Me₄NCl (0.1M or 0.01M). Data analysis for all tiration results was performed following the same procedures as previously for the determination of protonation constants [25] and stability constants [13] [38] using the computer program SCO 76 [39]. ¹H-NMR: Varian-A-60, Varian-EM-360A or Bruker-SY-200 spectrometer. ¹³C-NMR: Varian-XL-100 or Bruker-SY-200 spectrometer. Chemical shifts δ are given in ppm with tetramethyl-silane as internal standard. MS and microanalyses were performed by the 'Service de Spectrométrie de Masse' and by the 'Service de Microanalyse', resp., Institut de Chimie, Strasbourg.

2. Protected Diamines 7 and 14. The diamine, K_2CO_3 , and H_2O were heated to 80°. To the vigorously stirred soln., TsCl was added in batches within *ca*. 30 min. Stirring was continued at 80° for 24 h. Then the mixture was allowed to cool to r.t. After filtration, the white solid was washed with H_2O and the desired compound crystallized from hot EtOH.

1,7-Bis(p-tolylsulfonyl)-1,7-heptanediamine (= N,N'-(heptane-1,7-diyl)bis(p-toluenesulfonamide); 7). From 1,7-heptanediamine (10 g, 0.076 mol), K₂CO₃ (52 g), H₂O (1 l), and TsCl (30.74 g, 0.16 mol). Yield 32.66 g (97%), m.p. 144–145°. ¹H-NMR (CDCl₃): 1.22 (br., 10 H, CH₂CH₂CH₂); 2.42 (s, 2 CH₃); 2.85 (br., 2 CH₂N); 5.12 (br., 1 NH); 7.28, 7.78 (2*m*, 8 arom. H). ¹³C-NMR (CDCl₃): 143.4, 137.4, 129.8, 127.2 (arom. C); 43.2 (CH₂N); 29.3, 28.4, 26.2 (CH₂CH₂CH₂); 21.6 (CH₃). Anal. calc. for C₂₁H₃₀N₂O₄S₂ (438.58): C 57.50, H 6.89, N 6.38; found: C 57.64, H 6.98, N 6.39.

1,10-Bis(p-tolylsulfonyl)-1,10-decanediamine (= N,N⁷-(decane-1,10-diyl)bis(p-toluenesulfonamide); 14).From 1,10-decanediamine (25 g, 0.145 mol), K₂CO₃ (100 g), H₂O (1.5 l), and TsCl (55.32 g, 0.288 mol). Yield 66.2 g (95%), m.p. 127–128°. ¹H-NMR (CDCl₃): 1.15 (br., 16 H, CH₂CH₂CH₂); 2.35 (*s*, 2 CH₃); 2.90 (*m*, 2 CH₂N); 5.10 (br., 2 NH); 7.18, 7.69 (*m*, 8 arom. H). ¹³C-NMR (CDCl₃): 143.1, 137.0, 129.5, 127.0 (arom. C); 43.0 (CH₂N); 29.2, 28.9, 28.7, 26.2 (CH₂CH₂CH₂); 21.3 (CH₃). Anal. calc. for C₂₄H₃₆N₂O₄S₂ (480.66): C 59.96, H 7.54, N 5.82; found: C 59.84, H 7.47, N 5.99.

3. Dinitriles 8 and 15. To a mixture of compounds 7 or 14, K_2CO_3 , and DMF, a soln. of acrylonitrile in DMF was added dropwise at r.t. Stirring was continued for 15 h. After evaporation to dryness, the residue was taken up in CH_2Cl_2 and washed with H_2O and brine. The org. layer was dried over MgSO₄ and evaporated. The pure compounds 8 and 15 were obtained by crystallization from hot EtOH.

4,12-Bis(p-tolylsulfonyl)-4,12-diazapentadecane-1,15-dinitrile (= N,N'-bis(2-cyanoethyl)-N,N'-(heptane-1,7-diyl)bis(p-toluenesulfonamide); **8**). From **7** (27.19 g, 0.062 mol), K_2CO_3 (60 g), DMF (300 ml), acrylonitrile (10 ml, 0.15 mol), and DMF (20 ml). Yield 21.61 g (64%), m.p. 98–99°. ¹H-NMR (CDCl₃): 1.30 (br., 10 H, CH₂CH₂CH₂); 2.43 (s, 2 CH₃); 2.69 (m, 2 CH₂CN); 3.20 (m, 8 H, CH₂N); 7.30, 7.80 (m, 8 arom. H). ¹³C-NMR (CDCl₃): 144.1, 135.9, 130.1, 127.3 (arom. C); 119.8 (CN); 49.6, 44.5 (CH₂N); 28.5, 26.4 (CH₂CH₂CH₂CH₂); 21.6 (CH₃); 19.1 (CH₂CN). Anal. calc. for C₂₇H₃₆N₄O₄S₂ (544.64): C 59.53, H 6.66, N 10.29; found: C 59.68, H 6.67, N 10.26.

4,15-Bis(p-tolylsulfonyl)-4,15-diazaoctadecane-1,18-dinitrile (= N,N'-bis(2-cyanoethyl)-N,N'-(decane-1,10-diyl)bis(p-toluenesulfonamide); **15**). From **14** (30 g, 0.062 mol), K₂CO₃ (60 g), DMF (300 ml), acrylonitrile (10 ml, 0.15 mol), and DMF (20 ml). Yield 29.67 g (81%), m.p. 127-128°. ¹H-NMR (CDCl₃): 1.28 (br., 16 H, CH₂CH₂CH₂); 2.45 (s, 2 CH₃); 2.75 (m, 8 H, CH₂N); 3.28 (m, 2 CH₂CN); 7.30, 7.70 (m, 8 arom. H). ¹³C-NMR (CDCl₃): 144.0, 136.0, 130.1, 127.4 (arom. C); 117.7 (CN); 49.3, 44.4 (CH₂N); 29.3, 29.1, 28.7, 26.6 (CH₂CH₂CH₂); 21.6 (CH₃); 19.0 (CH₂CN). Anal. calc. for C₃₀H₄₂N₄O₄S₂ (586.78): C 61.40, H 7.21, N 9.54; found: C 61.44, H 7.16, N 9.59.

4. Diamines 9 and 16. Under Ar, 8 (10 g, 18 mmol) or 15 (10 g, 17 mmol) in 80 ml of THF were cooled and stirred in an ice bath. To this, $1 \le B_2 H_6$ in 150 ml of THF was added and the mixture refluxed overnight. Then, it was allowed to cool to r.t., and stirring was continued for another 6 h. The mixture was cooled in an ice bath, and 15 ml of H₂O were added cautiously. The solvant was evaporated; 6N HCl (300 ml) was added and the soln. refluxed overnight. After evaporation, the residue was partitioned between 1N NaOH (400 ml) and CH₂Cl₂ (300 ml). The org. layer was separated and the aq. layer extracted again with CH₂Cl₂ (2 × 100 ml). The combined org. layers were dried over MgSO₄ and evaporated to leave 9 (10 g) and 16 (10 g), resp., as a pale yellow oil. These compounds must be stored under N₂ and were converted to 10 or 17 without further purification.

4.12-Bis(p-tolylsulfonyl)-4.12-diazapentadecane-1.15-diamine (= N,N'-bis(3-aminopropyl-N,N'-(heptane-1,7-diyl)bis(p-toluenesulfonamide); 9). ¹H-NMR (CDCl₃): 1.29 (br., 10 H, CH₂CH₂CH₂); 1.57 (br., 8 H,

 $CH_2CH_2NH_2$); 2.43 (s, 2 CH₃); 2.72 (m, 2 CH₂NH₂); 3.13 (m, 8 H, CH₂NTs); 7.29, 7.80 (m, 8 arom. H). ¹³C-NMR (CDCl₃): 143.3, 137.1, 129.9, 127.3 (arom. C); 48.6, 46.1 (CH₂NTs); 39.3 (CH₂NH₂); 32.5, 28.7, 26.7 (CH₂CH₂CH₂); 21.6 (CH₃). Anal. calc. for $C_{27}H_{44}N_4O_4S_2$ (552.77): C 58.66, H 8.02, N 10.29; found: C 57.28, H 8.22, N 9.77.

4,15-Bis(p-tolylsulfonyl)-4,15-diazaoctadecane-1,18-diamine (= N,N'-bis(3-aminopropyl)-N,N'-(decane-1,10-diyl)bis(p-toluenesulfonamide); **16**). ¹H-NMR (CDCl₃): 1.25 (br., 16 H, CH₂CH₂CH₂); 1.68 (br., 8 H, CH₂CH₂NH₂); 2.40 (s, 2 CH₃); 2.72 (m, 2 CH₂NH₂); 3.15 (m, 8 H, CH₂NTs); 7.30, 7.70 (m, 8 arom. H). ¹³C-NMR (CDCl₃): 143.2, 137.1, 129.8, 127.3 (arom. C); 48.6, 46.0 (CH₂NTs); 39.2 (CH₂NH₂); 32.4, 29.5, 29.3, 28.8, 26.9 (CH₂CH₂CH₂); 21.6 (CH₃). Anal. calc. for C₃₀H₅₀N₄O₄S₂ (594.85): C 60.57, H 8.47, N 9.42; found: C 58.91, H 8.58, N 9.54.

5. Tetratoluenesulfonamides 10 and 17. To a stirred mixture of 9 or 16, Et₃N, and THF, TsCl was added at r.t. and stirring continued for 12-16 h. After evaporation, the yellow residue was taken up in CH₂Cl₂, washed with 1N HCl or 1N NaOH, H₂O or sat. NaCl soln., and dried over MgSO₄. Evaporation gave a colored residue which was purified by chromatography on alumina with CH₂Cl₂/0 to 1% MeOH.

N,N'-4,12-Tetrakis(p-tolylsulfonyl)-4,12-diazapentadecane-1,15-diamine (= N,N'-[4,12-bis(p-tolylsulfonyl)-4,12-diazapentadecan-1,15-diyl]bis(p-toluenesulfonamide); **10**). From **9** (5 g, 9 mmol), Et₃N (10 ml), THF (100 ml), and TsCl (3.6 g, 18 mmol); CH₂Cl₂ (300 ml), 1N HCl (250 ml), and H₂O (200 ml). Yield 6 g (85%). ¹H-NMR (CDCl₃): 1.21 (br., 10 H, CH₂CH₂CH₂); 1.75 (br., 2 CH₂CH₂NHTs); 2.40 (s, 4 CH₃); 3.05 (br., 12 H, CH₂N); 5.60 (br., 2 NH); 7.29, 7.65, 7.80 (m, 16 arom. H). ¹³C-NMR (CDCl₃): 143.5, 143.4, 137.2, 136.4, 129.9, 127.3 (arom. C); 48.9, 45.7, 40.2 (CH₂N); 29.1, 28.3, 26.3 (CH₂CH₂CH₂); 21.5 (CH₃). Anal. calc. for C₄₂H₅₈N₄O₈S₄·1 MeOH (907.19): C 56.47, H 6.77, N 6.27; found: C 56.15, H 6.60, N 6.44.

N,N',4,15-*Tetrakis*(p-tolylsulfonyl)-4,15-diazaoctadecane-1,18-diamine (= N,N'-[4,15-bis(p-tolylsulfonyl)-4,15-diazaoctadecan-1,18-diyl]bis(p-toluenesulfonamide); **17**). From **16** (10 g, 16.8 mmol), Et₃N (9.3 ml), THF (120 ml), and TsCl (8 g, 40 mmol); CH₂Cl₂ (300 ml), 1N HCl (300 ml), sat. NaCl (300 ml), 1N NaOH (300 ml), and H₂O (200 ml). Yield 12.75 g (84%). ¹H-NMR (CDCl₃): 1.19 (br., 16 H, CH₂CH₂CH₂); 1.65 (*m*, 2 CH₂CH₂NHTs); 2.35 (*s*, 4 CH₃); 3.05 (*m*, 12 H, CH₂N); 5.60 (br., 2 NH); 7.25, 7.65, 7.78 (*m*, 16 arom. H). ¹³C-NMR (CDCl₃): 143.5, 137.3, 136.5, 130.0, 127.3 (arom. C); 49.0, 45.8, 40.5 (CH₂N); 29.2, 26.8 (CH₂CH₂CH₂); 21.6 (CH₃). Anal. calc. for C₄₄H₆₂N₄O₈S₄ (903.20): C 58.50, H 6.92, N 6.20; found: C 58.54, H 7.03, N 6.38.

6. Diols 11 and 18. Compounds 7 or 14, K_2CO_3 , and DMF were stirred and heated to 80°. To this, a soln. of 3-chloro-1-propanol in DMF was added dropwise. Stirring at 80° was continued for 24 to 36 h. After evaporation, the residue was taken up in CH₂Cl₂, washed with H₂O, 1N NaOH, sat. NaCl soln., and dried over MgSO₄. Evaporation gave a colored oil which was purified by chromatography on alumina with CH₂Cl₂/2.5 to 4% MeOH.

4,12-Bis(p-toly/sulfonyl)-4,12-diazapentadecane-1,15-diol (= N,N'-bis(3-hydroxypropyl)-N,N'-(heptane-1,7-diyl)bis(p-toluenesulfonamide); **11**). From 7 (10 g, 0.023 mol), K₂CO₃ (15.75 g), DMF (150 ml), and 3-chloro-1-propanol (10.77 g, 0.11 mol) in DMF (30 mol); CH₂Cl₂ (400 ml), H₂O (500 ml), 1N NaOH (150 ml), and sat. NaCl soln. (150 ml). Yield 8.6 g (88%). ¹H-NMR (CDCl₃): 1.25 (br., 10 H, CH₂CH₂CH₂); 1.75 (m, 2 CH₂CH₂OH); 2.35 (s, 2 CH₃); 3.10 (m, 10 H, CH₂N, OH); 3.70 (br., 2 CH₂OH); 7.30, 7.80 (m, 8 arom. H). ¹³C-NMR (CDCl₃): 143.5, 136.8, 129.9, 127.2 (arom. C); 59.3 (CH₂OH); 48.9, 45.5 (CH₂N); 31.8, 28.7, 26.6 (CH₂CH₂CH₂); 21.6 (CH₃). Anal. ealc. for C₂₇H₄₂N₂O₆S₂ (554.74): C 58.45, H 7.63, N 5.05; found: C 58.56, H 7.64, N 5.18.

4,15-Bis(p-tolylsulfonyl)4,15-diazaoctadecane-1,18-diol (N,N'-bis(3-hydroxypropyl)-N,N'-(decane-1,10-diyl)bis(p-toluenesulfonamide); **18**). From **14** (10 g, 0.02 mol) K₂CO₃ (14.38 g), DMF (130 ml), and 3-chloro-1-propanol (9.84 g, 0.1 mol) in DMF (30 ml); CH₂Cl₂ (400 ml), H₂O (500 ml), and sat. NaCl soln. (100 ml). M.p. 84-86°, yield 9.0 g (73%). ¹H-NMR (CDCl₃): 1.20 (br., 16 H, CH₂CH₂CH₂); 1.75 (m, 2 CH₂CH₂OH); 2.35 (s, 2 CH₃); 3.32 (m, 14 H, CH₂N, CH₂OH, OH); 7.25, 7.69 (m, 8 arom. H). ¹³C-NMR (CDCl₃): 143.4, 136.9, 130.0, 127.3 (arom. C); 59.4 (CH₂OH); 48.9, 45.5 (CH₂N); 31.8, 29.2, 28.8, 26.8 (CH₂CH₂CH₂); 21.6 (CH₃). Anal. calc. for C₃₀H₄₈N₂O₆S₂ (568.82): C 60.37, H 8.10, N 4.69; found: C 60.25, H 8.19, N 4.70.

7. Dimesylates 12 and 19. Compound 11 or 18, Et_3N , and $dry CH_2Cl_2$ were stirred and cooled in an ice bath. To this was added MsCl. Stirring was continued for 30–90 min before the soln. was allowed to warm to r.t., and it was stirred for another 3–4 h. The mixture was washed with 1N HCl, 1N NaOH, and dried over MgSO₄. Evaporation gave an orange liquid which was used for the next step without further purification.

4,12-Bis(p-tolylsulfonyl)-4,12-diazapentadecane-1,15-diyl Bis(methanesulfonate) (12). From 11 (4.09 g, 7.3 mmol), Et₃N (5 ml), CH₂Cl₂ (120 ml), and MsCl (1.42 ml, 18.4 mmol); 1N HCl (2 × 50 ml) and 1N NaOH (50 ml). Yield 4.70 g (90%). ¹H-NMR (CDCl₃): 1.29 (br., 10 H, CH₂CH₂CH₂); 2.0 (m, 2 CH₂CH₂OMs); 2.41 (s, 2 CH₃C₆H₄); 3.08 (s, 2 CH₃SO₂); 3.15 (m, 8 H, CH₂N); 4.30 (m, 2 CH₂OMs); 7.30, 7.70 (m, 8 arom. H). ¹³C-NMR (CDCl₃): 143.7, 136.6, 130.1, 127.4 (arom. C); 68.1 (CH₂OMs); 49.2, 45.2 (CH₂N); 37.3 (CH₃SO₂); 29.0, 28.6, 26.6

 $(CH_2CH_2CH_2)$; 21.6 (CH₃). Anal. calc. for $C_{29}H_{46}N_2O_{10}S_4 \cdot \frac{1}{2}$ CH₂Cl₂ (753.37): C 47.02, H 6.28, N 3.71; found: C 47.34, H 6.60, N 3.85.

4,15-Bis(p-tolylsulfonyl)-4,15-diazaoctadecane-1,18-diyl Bis(methanesulfonate) (19). From 18 (5.5 g, 92 mmol), Et₃N (5 ml), CH₂Cl₂ (170 ml), and MsCl (1.8 ml, 23.3 mmol); 1N HCl (2×50 ml). Yield 6.59 g (95%). ¹H-NMR (CDCl₃): 1.25 (br., 16 H, CH₂CH₂CH₂); 2.0 (*m*, 2 CH₂CH₂OMs); 2.40 (*s*, 2 CH₃C₆H₄); 3.03 (*s*, 2 CH₃SO₂); 3.19 (*m*, 8 H, CH₂N); 4.29 (*m*, 2 CH₂OMs); 7.30, 7.70 (*m*, 8 arom. H). ¹³C-NMR (CDCl₃): 143.6, 136.6, 130.0, 127.3 (arom. C); 67.9 (CH₂OMs); 49.2, 45.0 (CH₂N); 37.3 (CH₃SO₂); 29.4, 29.2, 28.7, 26.8 (CH₂CH₂CH₂); 21.6 (CH₃C₆H₄). Anal. calc. for C₃₂H₅₂N₂O₁₀S₄ · ½ CH₂Cl₂ (752.99): C 51.12, H 6.99, N 3.67; found: C 50.29, H 7.33, N 4.02.

8. Protected Macrocycles **13** and **20**. 1,5,9,17,21,25-Hexakis(p-tolylsulfonyl)-1,5,9,17,21,25-hexaazacyclodotriacontane (**13**). A mixture of **10** (2.71 g, 3.1 mmol), Cs₂CO₃ (5.12 g), and DMF (100 ml) was stirred and heated to 100°. To this, a soln. of **12** (2.23 g, 3.1 mmol) in DMF (30 ml) was added dropwise within 15 min. Stirring at 100° was continued for 24 h. After evaporation, the yellow residue was taken up in CH₂Cl₂ (120 ml), washed with 1N NaOH (100 ml), 1N H₂SO₄ (2 × 100 ml), sat. NaCl soln. (100 ml), and dried over MgSO₄. Evaporation gave 6 g of mixture. Pure **13** (1.08 g, 25%) was obtained as a glass after chromatography on alumina (150 g) with CH₂Cl₂/ 0.5% MeOH, then on silica gel (100 g) with CH₂Cl₂/0 to 4% MeOH, and finally by gel permeation (HPLC, *Prepak*, *Waters*). ¹H-NMR (CDCl₃): 1.30 (br., 20 H, CH₂CH₂CH₂); 1.90 (br., 8 H, NCH₂CH₂CH₂N); 2.40 (*s*, 6 CH₃); 3.10 (br., 24 H, CH₂N); 7.30, 7.70 (*m*, 24 arom. H). ¹³C-NMR (CDCl₃): 143.5, 143.3, 137.3, 136.8, 129.8, 127.3 (arom. C); 49.2, 47.4, 46.7 (CH₂N); 29.3, 29.0, 26.6 (CH₂CH₂CH₂); 21.5 (CH₃). MS: 1378 (*M*⁺), 1223 (*M*⁺ - Ts), 1068 (*M*⁺ - 2Ts), 913 (*M*⁺ - 3Ts), 758 (*M*⁺ - 4Ts), 603 (*M*⁺ - 5Ts). Anal. calc. for C₆₈H₉₄N₆O₁₂S₆ (1379.83): C 59.18, H 6.86, N 6.09; found: C 59.08, H 6.99, N 6.02.

1,5,9,20,24,28-Hexakis(p-tolylsulfonyl)-1,5,9,20,24,28-hexaazacyclooctatriacontane (20). A mixture of 17 (8.14 g, 9 mmol), Cs₂CO₃ (17.59 g), and DMF (120 ml) was stirred and heated to 90°. To this, a soln. of 19 (6.50 g, 9 mmol) in DMF (30 ml) was added dropwise within 10 min. Stirring at 90° was continued for 48 h. After evaporation, the colored residue was taken up in CH₂Cl₂ (250 ml), washing with 1N NaOH (200 ml) produced an emulsion which was broken by addition of sat. NaCl soln. (100 ml). The org. layer was dried over MgSO₄. Evaporation gave 12.96 g of a mixture. Pure 20 (2.39 g, 25%) was obtained as a glass after chromatography on alumina (500 g) with CH₂Cl₂/0.5% MeOH, then on silica gel (200 g) with CH₂Cl₂/2% MeOH, and finally by gel permeation (HPLC, *Prepak, Waters*). ¹H-NMR (CDCl₃): 1.27 (br., 32 H, CH₂CH₂CH₂); 1.85 (br., 8 H, NCH₂CH₂CH₂N); 2.42 (s, 6 CH₃); 3.13 (br., 24 H, CH₂N); 7.30, 7.70 (m, 24 arom. H). ¹³C-NMR (CDCl₃): 1434, 1366, 129, 9, 127.3 (arom. C); 49.1, 47.4, 46.5 (CH₂N); 29.5, 29.3, 26.8 (CH₂CH₂CH₂); 21.6 (CH₃). MS: 1462 (M⁺), 1307 (M⁺ - Ts), 1152 (M⁺ - Ts), 997 (M⁺ - 3Ts). Anal. calc. for C₇₄H₁₀N₆O₁₂S₆ (1463.99): C 60.70, H 7.29, N 5.74; found: C 60.58, H 7.32, N 5.84.

9. Deprotection of Macrocycles 13 and 20. Compound 13 or 20, phenol, and a 33% soln. of HBr in AcOH (60 ml) were refluxed for 16–18 h under a well ventilated hood. Evaporation gave $1 \cdot 6$ HBr or $2 \cdot 6$ HBr which was suspended in Et₂O (50 ml), filtered, and washed with Et₂O (150 ml). The crude hexahydrobromides were dissolved in H₂O (15–25 ml) and passed over *Dowex 1 × 8* (basic form). Amine 2 is insoluble in H₂O; it was, therefore, eluted with H₂O/EtOH 1:1. The soln. containing 1 and 2 were acidified to pH 2 with conc. HCl and the solvent removed by evaporation. The residue was precipitated from aq. EtOH giving $1 \cdot 6$ HCl or $2 \cdot 6$ HCl.

1,5,9,17,21,25-Hexaazacyclodotriacontane (1). From 13 (1.11 g, 0.8 mmol) and phenol (2.5 g). Yield 0.48 g (92%), m.p. > 260°. ¹H-NMR (D₂O): 1.29 (br., 12 H, CH₂CH₂CH₂); 1.59 (br., 8 H, CH₂CH₂N); 2.02 (br., 8 H, NCH₂CH₂CH₂N); 3.06 (m, 24 H, CH₂N). ¹³C-NMR (D₂O): 48.8, 45.9, 45.5 (CH₂N); 28.4, 26.3, 26.2, 23.6 (CH₂CH₂CH₂). Anal. calc. for C₂₄H₆₄Cl₆N₆ (673.51): C 45.15, H 9.62, N 12.15; found: C 45.07, H 9.58, N 12.22.

1,5,9,20,24,28-Hexaazacyclooctatriacontane (2). From 20 (1.30 g, 0.88 mmol) and phenol (2.5 g). Yield 0.62 g (93%), m.p. > 260°. ¹H-NMR (D₂O): 1.38 (br., 24 H, CH₂CH₂CH₂); 1.74 (br., 8 H, CH₂CH₂N); 2.17 (br., 8 H, NCH₂CH₂CH₂N); 3.22 (br., 24 H, CH₂N). ¹³C-NMR (D₂O): 49.1, 46.0, 45.6 (CH₂N); 29.2, 29.0, 26.7, 26.6, 23.9 (CH₂CH₂CH₂). Anal. calc. for C₃₂H₇₆Cl₆N₆ (757.67): C 50.72, H 10.11, N 11.09; found: C 50.56, H 10.22, N 11.02.

10. 4,15-Diazaoctadecane-1,18-diamine (5). Under a well ventilated hood, 17 (2.40 g, 2.65 mmol), phenol (2.3 g), and a 33 % soln. of HBr in AcOH (150 ml) were refluxed for 16 h. The soln. was allowed to cool and the solid decanted, filtered, washed with Et₂O (2×50 ml), dissolved in H₂O (30 ml), and passed over *Dowex 1 × 8* (basic form). The soln. containing **5** was acidified with conc. HCl to pH 2 and the solvent evaporated. The residue was precipitated from aq. EtOH giving **5** ·4 HCl (1.1 g, 95%), m.p. > 260°. ¹H-NMR (D₂O): 1.41 (br., 12 H, CH₂CH₂CH₂); 1.77 (br., 2 CH₂CH₂M₂⁺-); 2.12 (br., 2 NCH₂CH₂CH₂N); 3.28 (br., 12 H, CH₂N). ¹³C-NMR (D₂O): 49.4, 45.9 (CH₂NH₂⁺-); 38.15 (CH₂NH₃⁺); 29.9, 29.7, 27.2, 27.0, 25.2 (CH₂CH₂CH₂). Anal. calc. for C₁₆H₄₂Cl₄N₄ · 2 H₂O (466.34): C 41.20, H 9.51, N 12.01; found: C 41.28, H 9,47, N 12.16.

11. Decane-1,10-diyl Bis(methanesulfonate) (21). At r.t., 1,10-decanediol (11.7 g, 0.067 mol), Et₃N (47 ml), dry CH₂Cl₂ (200 ml), and dry THF (50 ml) were stirred. To this, MsCl (16 ml) in dry CH₂Cl₂ (130 ml) was added dropwise within 1 h. Stirring at r.t. was continued for 15 h. After filtration, the soln. was washed with 10% HCl (150 ml), sat. NaHCO₃ soln. (200 ml), and H₂O (200 ml), dried over MgSO₄, and evaporated. Pure 21 (20.62 g, 93%) was obtained by crystallization from CH₂Cl₂/hexane, m.p. 74–76°. ¹H-NMR (CDCl₃): 1.40 (br., 12 H, CH₂CH₂CH₂); 1.72 (br., 2 CH₂CH₂OMs); 3.10 (s, 2 CH₃); 4.25 (m, 2 CH₂OMs). ¹³C-NMR (CDCl₃): 69.8 (CH₂OMs); 37.6 (CH₃); 29.4 (CH₂CH₂OMs); 29.3, 29.1, 25.6 (CH₂CH₂CH₂). Anal. calc. for C₁₂H₂₆O₆S₂ (330.45): C 43.61, H 7.93; found: C 43.67, H 7.93.

12. N,N',4,8,19,23-Hexakis(p-tolylsulfonyl)-4,8,19,23-tetraazahexaicosane-1,26-diamine (= N,N'-[4,8,19,23-tetrakis(p-tolylsulfonyl)-4,8,19,23-tetraazahexaicosan-1,26-diyl]bis(p-toluenesulfonamide); **22**). Na (1.74 g) and dry MeOH (100 ml) were stirred at r.t. under Ar until dissolution. To this, 17.96 g of N,N',4-tris(p-tolylsulfonyl)-4-azaheptanediamine (17.96 g) prepared as described in [25] was added and the mixture refluxed for 1 h. After evaporation, the disodium salt was dissolved in DMF (100 ml), stirred, and heated to 90°. To this, a soln. of compound **21** (2 g) in DMF (100 ml) was added within 14 h. Stirring at 90° was continued for 10 h. After filtration, the solvent was removed, and the yellow residue was taken up in CH₂Cl₂ (100 ml), washed with N NaOH (100 ml), H₂O (100 ml), and sat. NaCl soln. (100 ml), and dried over MgSO₄. Pure **22** (0.53 g, 6%) was obtained after chromatography on alumina with CH₂Cl₂. ¹H-NMR (CDCl₃): 1.25 (br., 12 H, CH₂CH₂CH₂); 1.75 (br., 12 H, CH₂CH₂N); 2.40 (*s*, 6 CH₃); 3.10 (br., 20 H, CH₂N); 5.30 (br., 2 NH); 7.25, 7.68, 7.80 (*m*, 24 arom. H). ¹³C-NMR (CDCl₃): 143.3, 143.2, 129.8, 127.1 (arom. C); 49.0, 47.3, 46.4, 46.2, 40.0 (CH₂N); 29.3, 29.2, 29.1, 28.8, 28.6, 26.7 (CH₂CH₂CH₂); 21.5 (CH₃). Anal. calc. for C₆₄H₈₈N₆O₁₂S₆ (1325.74): C 57.97, H 6.69, N 6.34; found: C 56.63, H 7.25, N 6.05.

13. 4,8,19,23-Tetraazahexaicosane-1,26-diamine (6). Under a well ventilated hood, **22** (0.52 g), phenol (1.50 g), and a 33% soln. of HBr in AcOH (100 ml) were refluxed for 16 h. The soln. was allowed to cool to r.t. and the solid decanted, filtered, washed with Et₂O (2×50 ml), dissolved in H₂O (20 ml), and passed over *Dowex 1 × 8* (basic form). The soln. containing **6** was acidified to pH 2 with conc. HCl and the solvent evaporated. The residue was precipitated from aq. EtOH giving **6** · 6 HCl (0.22 g, 93%), m.p. > 260°. ¹H-NMR (D₂O): 1.37 (br., 12 H, CH₂CH₂CH₂); 1.74 (br., 2 CH₂CH₂NH₃⁺); 2.06 (br., 8 H, CH₂CH₂NH₂⁺-); 3.09 (m, 20 H, CH₂N). ¹³C-NMR (D₂O): 49.5, 46.3, 45.8 (CH₂N); 30.0, 29.8, 27.3, 27.1, 25.3, 24.2 (CH₂CH₂CH₂). Anal. calc. for C₂₂H₅₈Cl₆N₆· 2 EtOH (711.56): C 43.88, H 9.91; found: C 44.24, H 9.90.

14. N,N',3-Tris(p-tolylsulfonyl)-3-azapentanediamine (= N,N'-[3-(p-tolylsulfonyl)-3-azapentane-1,5-diyl]bis(p-toluenesulfonamide); **23**) was prepared by following the procedure described in [33].

15. Bissodium Salt 24 of 23. Na (1.42 g) and dry MeOH (200 ml) were stirred at r.t. under Ar until dissolution. To this, 23 (16.43 g, 0.029 mol) was added and the mixture refluxed for 4 h. After evaporation, the white solid (17.53 g) was dried under vacuum for 12 h.

16. 9-[(2H-Tetrahydropyran-2-yl)oxy]nonan-1-ol (25). Nonane-1,9-diol (60 g, 0.37 mol), conc. HCl (10 drops), and THF (20 ml) were stirred at r.t. To this, a soln. of 2 *H*-dihydropyran (10.5 g, 0.12 mol) in THF (60 ml) was added dropwise and stirring continued for 7 h. After evaporation, the residue was taken up in toluene (300 ml) and heated to 60° until homogeneous. The unreacted diol crystallized from soln. upon cooling to 4°. The solid was filtered, washed with toluene, and the combined filtrates were washed with H₂O (200 ml), dried over MgSO₄, and evaporated. Pure **25** (34 g) was obtained as an oil, after chromatography on silica gel with AcOEt/hexane 4:6. ¹H-NMR (CDCl₃): 1.50 (br., 20 H, CH₂CH₂CH₂); 3.60 (*m*, 7 H, CH₂OH, OH); 4.60 (br., OCHO). ¹³C-NMR (CDCl₃): 98.7 (OCHO); 67.5, 62.4, 62.1 (CH₂O); 32.6, 30.6 (CH₂CH₂O); 29.3, 26.1, 25.4, 19.5 (CH₂CH₂CH₂). Anal. calc. for C₁₄H₂₈O₃ (224.36): C 68.80, H 11.54; found: C 68.12, H 10.97.

17. 9-[(2H-Tetrahydropyran-2-yl)oxy]nonyl Methanesulfonate (26). A mixture of 25 (5.86 g, 0.024 mol), Et₃N (5 ml), and dry CH₂Cl₂ (60 ml) was stirred and cooled to -18° . To this, a soln. of MsCl (2.04 ml) in dry CH₂Cl₂ (20 ml) was added dropwise within 20 min. The mixture was allowed to warm to r.t. and stirring continued for 2 h. The soln. was washed with ice-water (50 ml), cold 1N HCl (50 ml), sat. NaHCO₃ soln. (50 ml), sat. NaCl soln. (50 ml), and dried over MgSO₄. Evaporation left 26 (98%) as an oil which was used without further purification for the following step. ¹H-NMR (CDCl₃): 1.50 (br., 20 H, CH₂CH₂CH₂); 3.05 (*s*, CH₃); 3.60 (*m*, 2 CH₂O); 4.25 (*t*, CH₂OMs). ¹³C-NMR (CDCl₃): 98.7 (OCHO); 70.1, 67.4, 62.1 (CH₂O); 37.1 (CH₃); 30.7, 29.6, 29.2, 28.9, 26.1, 25.4, 25.2, 19.5 (CH₂CH₂CH₂).

18. 10,13,16-Tris(p-tolylsulfonyl)-1,25-bis[(2H-tetrahydropyran-2-yl)oxy]-10,13,16-triazapentaicosane (27). Under Ar, 24 (17.53 g, 28.7 mmol) and dry DMF (300 ml) were stirred and heated to 95°. To this, a soln. of 26

(18.35 g, 56.9 mmol) in dry DMF (150 ml) was added dropwise within 20 min. Stirring was continued at 95° for 6 h. After evaporation, the residue was taken up in CH₂Cl₂ (300 ml), washed with H₂O (300 ml), and dried over MgSO₄. Evaporation gave 25 g of a yellow liquid which was purified by chromatography on alumina (400 g) with CH₂Cl₂(0.5–1 % MeOH: 22 g (72 %) of **27**. ¹H-NMR (CDCl₃): 1.50 (br., 40 H, CH₂CH₂CH₂); 2.45 (*s*, 3 CH₃); 3.30 (*m*, 20 H, CH₂O, CH₂N); 4.55 (br., 2 OCHO); 7.30, 7.80 (*m*, 12 arom. H). ¹³C-NMR (CDCl₃): 143.5, 136.0, 130.1, 129.9, 127.5 (arom. C); 67.8 (OCHO); 62.5 (CH₂O); 50.2, 49.9, 48.1 (CH₂N); 31.0, 29.9, 29.7, 29.6, 29.4, 28.8, 26.8, 26.4, 25.7 (CH₂CH₂CH₂); 21.5 (CH₃); 19.9 (CH₂CH₂CH₂). Anal. calc. for C₅₃H₈₃N₃O₁₀S₃ (1018.39): C 62.50, H 8.21, N 4.12; found: C 62.44, H 8.45, N 4.43.

19. 10,13,16-Tris(p-tolylsulfonyl)-10,13,16-triazapentaicosane-1,25-diol (**28**). For 8 h, **27** (20.64 g, 20.2 mmol) and TsOH (3.85 g) were refluxed in H₂O/EtOH 5:95 (250 ml). The mixture was allowed to cool to r.t. and stirring continued for another 12 h. After evaporation, the residue was taken up in CH₂Cl₂ (300 ml) and washed with H₂O (200 ml). The aq. layer was further extracted with CH₂Cl₂ (2 × 100 ml). The org. layers were combined, dried over MgSO₄, and evaporated: 18 go fa mixture. Pure **28** (12 g, 70%) was obtained after chromatography on alumina (500 g) with CH₂Cl₂/3-5% MeOH and recrystallized from CH₂Cl₂/EtOH, m.p. 74–77°. ¹H-NMR (CDCl₃): 1.30 (br., 28 H, CH₂CH₂CH₂); 2.55 (s, 11 H, CH₃, OH); 3.45 (m, 16 H, CH₂N, CH₂OH); 7.30, 7.80 (m, 12 arom. H). ¹³C-NMR (CDCl₃): 143.5, 136.0, 130.0, 129.9, 127.3 (arom. C); 63.2 (CH₂OH); 50.2, 49.9, 48.2 (CH₂N); 32.9, 29.5, 29.4, 29.2, 28.8, 26.7, 25.8 (CH₂CH₂CH₂); 2.1.6 (CH₃). Anal. calc. for C4₃H₆rN₃O₆S₃ (850.16): C 60.74, H 7.94, N 4.94; found: C 60.45, H 8.00, N 4.98.

20. 10,13,16-Tris(p-tolylsulfonyl)-10,13,16-triazapentaicosane-1,25-diyl Bis(methanesulfonate) (29). At -18° , 28 (10.88 g, 12.8 mmol) and Et₃N (9 ml) were stirred in dry CH₂Cl₂ (150 ml). To this, a soln. of MsCl (2.2 ml) in dry CH₂Cl₂ (50 ml) was added dropwise. The mixture was allowed to warm to r.t., and stirring was continued for 18 h. The colored soln. was washed with ice-water (100 ml), cold 1N HCl (100 ml), sat. NaHCO₃ soln. (100 ml), and dried over MgSO₄. Evaporation gave 29 (12.6 g) as an oil which was used without further purification for the following step. ¹H-NMR (CDCl₃): 1.30 (br., 28 H, CH₂CH₂CH₂); 2.45 (s, 3 CH₃C₆H₄); 3.2 (m, s, 18 H, CH₂N, CH₃SO₂); 4.22 (t, 2 CH₂OMs); 7.30, 7.80 (m, 12 arom. H). ¹³C-NMR (CDCl₃): 134.6, 136.0, 130.1, 130.0, 127.5 (arom. C); 70.3 (CH₂OMs); 50.2, 50.0, 48.2 (CH₂N); 37.6 (CH₃SO₂); 29.4, 29.3, 29.2, 29.0, 28.7, 26.7, 25.5 (CH₂CH₂CH₂); 2.1.7 (CH₃C₆H₄). Anal. calc. for C₄₅H₇I_N3O₁₂S₅ (1006.34): C 53.70, H 7.11, N 4.17; found: C 53.79, H 7.19, N 4.03.

21. 1,4.7,17,20,23-Hexakis (p-tolylsulfonyl)-1,4.7,17,20,23-hexaazacyclodotriacontane (**30**). A mixture of **29** (7.02 g, 12.4 mmol), Cs₂CO₃ (24.24 g), and DMF (600 ml) was stirred and heated to 90°. To this, a soln. of **23** (12.50 g, 12.4 mmol) in DMF (250 ml) was added dropwise within 1 h. Stirring at 90° was continued for 71 h. The mixture was allowed to cool to r.t. and the solid removed by filtration. After evaporation, the residue was taken up in CHCl₃ (400 ml), washed with 1 N NaOH (200 ml), sat. NaCl soln. (200 ml), and dried over MgSO₄. Evaporation gave 20 g of mixture which was purified by chromatography on alumina (300 g) with CHCl₃. Pure **30** (8.18 g, 47%) was obtained by recrystallization from CH₂Cl₂/EtOH, m.p. 150-152°. ¹H-NMR (CDCl₃): 1.3 (br., 28 H, CH₂CH₂CH₂); 2.45 (s, 6 CH₃); 3.2 (br., 24 H, CH₂N); 7.30, 7.80 (m, 24 arom. H). ¹³C-NMR (CDCl₃): 143.5, 136.2, 135.7, 130.1, 127.6 (arom. C); 50.4, 50.2, 48.5 (CH₂N); 29.1, 28.8, 26.6 (CH₂CH₂CH₂); 21.7 (CH₃). MS: 1379 (M⁺ - TS), 1068 (M⁺ - 2TS), 913 (M⁺ - 3TS), 758 (M⁺ - 4TS). Anal. calc. for C₆₈H₉₄N₆O₁₂S₆ (1379.83): C 58.96, H 7.07, N 5.89; found: C 58.63, H 6.77, N 5.96.

22. 1,4,7,17,20,23-Hexaazacyclodotriacontane (3). A mixture of **30** (3.43 g, 2.48 mmol), phenol (3.2 g), and a 33% soln. of HBr in AcOH (200 ml) was refluxed for 46 h. The mixture was allowed to cool to r.t. and the solid decanted, filtered, washed with Et₂O (3 × 100 ml), and dissolved in H₂O and passed over *Dowex 1* × 8 (basic form). Since **3** is insoluble in H₂O, the resin was eluted with H₂O/MeOH 3:1. The soln. containing **3** was acidified with conc. HCl to pH 3, and after evaporation, $3 \cdot 6$ HCl (1.50 g, 90%) was recrystallized from H₂O/MeOH/EtOH, m.p. > 260°. ¹H-NMR (D₂O): 1.40 (br., 20 H, CH₂CH₂CH₂); 3.20 (*m*, 8 H, CH₂CH₂N); 3.55 (br., 24 H, CH₂N). ¹³C-NMR (D₂O): 49.6, 45.0, 44.3 (CH₂N); 29.0, 26.5 (CH₂CH₂CH₂CH₂). Anal. calc. for C₂₆H₆₄Cl₆N₆ (673.53): C 46.25, H 9.57, N 12.47; found: C 46.24, H 9.57, N 12.36.

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